

# Avian Influenza Type A (H5) Primers and Probe Set

## Targets Influenza A Clade 2.3.4.4b

IDT consumables (kit 10020786 contents)

Component	Quantity (µL)	Storage (°C)	Content
InfA (H5) Primer/probe mix (200 rxns)	250	-20	20X primer/probe stock solution targeting InfA H5 (FAM)
InfA (H5) PCR control assay (200 rxns)	250		20X Primer/Probe stock solution targeting InfA PCR control (SUN)  InfA (H5) PCR synthetic control (5 × 10 <sup>3</sup> copies/µL)

Additional reagents (not included)

Component	Quantity	Manufacturer	Catalog #
PrimeTime 1-Step 4X Broad-Range Master Mix	1 mL	IDT	10011744
Nuclease Free Water	10 x 2 mL	IDT	11-04-02-01
InfA (H5) positive sample control	250 µL (1 × 10 <sup>4</sup> copies/µL)	IDT	10020808
Mycoplasma BVD Milk Extraction Kit	—	Udder Health Systems*	EXT-BVD1

\*<https://www.udderhealth.com>

## Milk sample lysis

- For each sample, combine 49.5 µL of the lysis buffer and 5.5 µL of the enzyme mix in a clean 1.5 mL tube. Vortex and centrifuge for 5 minutes at 16300 RCF.
- Dispense 50 µL into a 200 clear strip tube for each sample.
- Add 25 µL of milk for each sample to the appropriate tube, capping the tubes as you go.
- Vortex and spin down the tubes to collect the liquid.
- Heat the tubes to 60 °C for 30 minutes.
- Heat the tubes to 95 °C for 15 minutes.
- Allow the tubes to cool to 60 °C or below.
- Spin the tubes down for 5 minutes at 16300 RCF to pellet the solid material.
- Use 5 µL of the aqueous layer as sample in a 20 µL reaction. For best results, prepare fresh samples for each reaction rather than storing the crude lysates.

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Table 1. Positive control RT-qPCR setup.

Component	Volume (μL)
InfA (H5) Primer/probe mix	1
InfA (H5) PCR control	1
PrimeTime 1-Step 4X Broad-Range Master Mix	5
InfA (H5) positive sample control	1
Nuclease-free water	12
<b>Total</b>	<b>20</b>

Table 2. Crude lysate RT-qPCR setup

Component	Volume (μL)
InfA (H5) Primer/probe mix	1
InfA (H5) PCR control	1
PrimeTime 1-Step 4X Broad-Range Master Mix	5
Crude lysis sample	5
Nuclease-free water	8
<b>Total</b>	<b>20</b>

Table 3. RT-qPCR cycling protocol.

Step	Cycles	Temperature (°C)	Time
Reverse transcription	1	50	15 min
Enzyme activation	1	95	3 min
Denaturation	40	95	15 sec
Annealing/extension	40	60	1 min

## Run RT-PCR

Place the plate in the real-time PCR instrument and start the cycling program.

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