

# VARIANT*Plex* -HT Comprehensive Tissue & Blood

## Description

The VARIANT*Plex* Comprehensive Tissue & Blood panel is a balanced pool of gene-specific primer (GSP) oligonucleotides that is optimized for use with VARIANT*Plex*<sup>™</sup> reagents and molecular barcode (MBC) adapters to produce targeted NGS libraries. This product insert should be used in conjunction with VARIANT*Plex* -HT protocol for Illumina® (RA-DOC-058).

VARIANT*Plex* Comprehensive Tissue & Blood contains **26,535** GSPs targeting **460** genes commonly mutated in solid tumor and blood cancer types as well as microsatellite instability (**MSI**), tumor mutational burden (**TMB**), and homologous recombination deficiency (**HRD**) status.

Description	Part number	Storage
VARIANT <i>Plex</i> Comprehensive Tissue & Blood GSP1 - 8 reactions	SA24315081	-20°C ± 10°C
VARIANT <i>Plex</i> Comprehensive Tissue & Blood GSP2 - 8 reactions	SA24315082	

## Required reagent volumes

Protocol reference	Protocol step	Reagent	Volume per reaction (µL)
A	Ligation Step 2 Elution	5mM NaOH	20
B	First PCR	VARIANT <i>Plex</i> Comprehensive Tissue & Blood GSP1	8
C	First PCR	10mM Tris-HCl pH 8.0	18
D	First PCR	Purified PCR1 eluate	16
E	Second PCR	VARIANT <i>Plex</i> Comprehensive Tissue & Blood GSP2	8

## Recommended PCR cycling

	Step	Temperature (°C)	Time	Cycles
First PCR reaction	1	95	3 min	1
	2	95	30 sec	
	3	58	10 sec	15
	4	60	10 min (100% ramp rate)	
	5	72	3 min	1
	6	4	Hold	1
Second PCR reaction	1	95	3 min	1
	2	95	30 sec	
	3	58	10 sec	20†
	4	65	10 min (100% ramp rate)	
	5	72	3 min	1
	6	4	Hold	1

†The number of PCR2 cycles may be decreased if you regularly experience library yields greater than 200 nM.

## Recommended reads and multiplexing

VARIANT*Plex* Comprehensive Tissue & Blood libraries should be sequenced to a minimum of **61M** reads. Starting read depth recommendations for standard profiling may be adjusted based on user needs.

## Archer™ Analysis settings

Sequencing data should be processed using Archer Analysis (v7.4, or greater). The VARIANT*Plex* Comprehensive Tissue & Blood panel is compatible with the **SNV/Indel, Copy Number Variation, CNV 2.0, ASCN, Structural Variation, MSI, TMB, and HRD** pipelines, found under the **DNA** Input Type. The recommended TMB settings in Archer Analysis are **High Threshold for TMB = 17.5** and **Low Threshold for TMB = 6** for VARIANT*Plex*

Comprehensive Tissue & Blood. As with most Analysis settings, these may be adjusted based on user needs. Selection of the DNA Target Coverage pipeline is optional (see the Archer Analysis User Guide for more details on setting up your analysis).

Archer Analysis processing of VARIANT<sup>™</sup>Pl<sup>™</sup>ex Comprehensive Tissue & Blood libraries requires a one-time upload of the Panel GTF. When performing DNA Target Coverage analysis, users must also select a Region of Interest BED file. Users may optionally add a Targeted Mutations VCF file for targeted SNV/Indel detection. Files can be obtained by contacting [archer-tech@idtdna.com](mailto:archer-tech@idtdna.com)

## Assay targets

### Integrative genomic biomarkers

MSI, TMB, HRD

### Genes

The following genes in the panel have full coding sequence coverage for SNV/Indel analysis. Coding sequence was determined using NCBI RefSeq transcripts<sup>†</sup>. These genes are also enabled for Copy Number Variation (CNV) analysis.

<i>ABL1</i>	<i>CBLB</i>	<i>DMD</i>	<i>FGFR3</i>	<i>IRF6</i>	<i>MLL2</i>	<i>PIK3CG</i>	<i>RHOA</i>	<i>SYK</i>
<i>ABL2</i>	<i>CBLC</i>	<i>DNMT3A</i>	<i>FGFR4</i>	<i>IRS2</i>	<i>MPL</i>	<i>PIK3R1*</i>	<i>RICTOR</i>	<i>TAF1</i>
<i>ACVR1</i>	<i>CCND1</i>	<i>DOT1L</i>	<i>FH</i>	<i>JAK1</i>	<i>MRE11A</i>	<i>PIK3R2</i>	<i>RIT1</i>	<i>TBL1XR1</i>
<i>ACVR2A</i>	<i>CCND2</i>	<i>EEF1A1</i>	<i>FLCN</i>	<i>JAK2</i>	<i>MSH2</i>	<i>PIK3R3</i>	<i>RNF43</i>	<i>TBX3</i>
<i>AKT1</i>	<i>CCND3</i>	<i>EEF2</i>	<i>FLNA</i>	<i>JAK3</i>	<i>MSH3</i>	<i>PIM1</i>	<i>ROS1</i>	<i>TCEB1</i>
<i>AKT2</i>	<i>CCNE1</i>	<i>EGFR</i>	<i>FLT1</i>	<i>JUN</i>	<i>MSH6</i>	<i>PLCB4</i>	<i>RPL5</i>	<i>TCF3</i>
<i>AKT3</i>	<i>CD28</i>	<i>EGR3</i>	<i>FLT3</i>	<i>KANSL1</i>	<i>MST1R</i>	<i>PLCG1</i>	<i>RPS6KB1</i>	<i>TCF7L2</i>
<i>ALK</i>	<i>CD70</i>	<i>EIF1AX</i>	<i>FLT4</i>	<i>KDM5A</i>	<i>MTOR</i>	<i>PLCG2</i>	<i>RPS14</i>	<i>TERT</i>
<i>ANKRD26</i>	<i>CD79A</i>	<i>ELF3</i>	<i>FOXA1</i>	<i>KDM5C</i>	<i>MUC6</i>	<i>PLXNB2</i>	<i>RPTOR</i>	<i>TET1</i>
<i>APC</i>	<i>CD79B</i>	<i>EP300</i>	<i>FOXA2</i>	<i>KDM6A</i>	<i>MUC16</i>	<i>PMS1</i>	<i>RRAS2</i>	<i>TET2</i>
<i>AR</i>	<i>CD274 (PD-L1)</i>	<i>EPAS1</i>	<i>FOXL2</i>	<i>KDR</i>	<i>MUTYH</i>	<i>PMS2</i>	<i>RUNX1</i>	<i>TFRC</i>
<i>ARAF</i>	<i>CDC25C</i>	<i>EPCAM</i>	<i>FOXO1</i>	<i>KEAP1</i>	<i>MYC</i>	<i>POLD1</i>	<i>RUNX1T1</i>	<i>TGFBR1</i>
<i>ARHGAP35</i>	<i>CDC73</i>	<i>EPHA2</i>	<i>FOXP1</i>	<i>KEL</i>	<i>MYCL (MYCL1)</i>	<i>POLE</i>	<i>RXRA</i>	<i>TGFBR2</i>
<i>ARID1A</i>	<i>CDH1</i>	<i>EPHA3</i>	<i>FOXQ1</i>	<i>KIF1A</i>	<i>MYCN</i>	<i>POLQ</i>	<i>SAMD9</i>	<i>TGIF1</i>
<i>ARID1B</i>	<i>CDK4</i>	<i>EPHA7</i>	<i>FUBP1</i>	<i>KIT*</i>	<i>MYD88</i>	<i>POLRMT</i>	<i>SAMD9L</i>	<i>THRAP3</i>

ARID2	CDK6	EPHB1	GATA1	KLF2	MYH9	PPARG	SCAF4	TLR4
ARID5B	CDK8	ERBB2	GATA2	KLF4	NBN	PPM1D	SDHA	TMSB4X
ASXL1	CDK12	ERBB3	GATA3	KLF5	NCOR1	PPP2R1A	SDHB	TNFAIP3
ASXL2	CDKN1A	ERBB4	GEN1	KMT2A (MLL)	NF1	PPP2R2A	SDHC	TNFRSF14
ATM*	CDKN1B	ERCC1	GLI1	KMT2B	NF2	PPP6C	SDHD	TP53
ATR	CDKN2A	ERCC2	GNA11	KMT2C	NFE2L2	PRDM1	SETBP1	TP63
ATRX	CDKN2B	ERCC4	GNA13	KMT2D (MLL2)	NFKBIA	PRKAR1A	SETD2	TRAF2
AURKA	CDKN2C	ERG	GNAQ	KRAS	NFKBIE	PRKD1	SF3B1	TRAF3
AURKB	CEBPA	ERRFI1	GNAS	KRT222	NIPBL	PRKDC	SH2B3	TRAF7
AXIN1	CHD1	ESR1	GPS2	LAMP1	NKX2-1	PSIP1	SH2D1A	TSC1
AXIN2	CHD3	ETNK1	GRIN2D	LATS1	NOTCH1*	PTCH1	SLC29A1	TSC2
AXL	CHD4	ETV6	GRM3	LATS2	NOTCH2	PTEN	SLX4	TSHR
B2M	CHD8	EWSR1	H3F3A	LEMD2	NOTCH3	PTMA	SMAD2	TXNIP
BAP1	CHEK1	EZH2	H3F3B	LRP1B	NOTCH4	PTPDC1	SMAD3	U2AF1
BARD1	CHEK2*	FAM46C	H3F3C	LUC7L2	NPM1	PTPN11	SMAD4	U2AF2
BCL2	CIC	FAM46D	HGF	LZTR1	NRAS	PTPRC	SMARCA4	UBA1
BCL2L1	COL5A1	FAM175A	HIST1H1C	MACF1	NSD1	PTPRD	SMARCB1	UNCX
BCL2L11	CREBBP	FANCA	HIST1H1E	MAP2K1 (MEK1)	NTRK1	PTPRT	SMC1A	USP9X
BCL6	CRKL	FANCC	HIST1H2BD	MAP2K2 (MEK2)	NTRK2	RAC1	SMC3	VHL
BCOR	CSDE1	FANCD2	HIST1H3B	MAP2K4	NTRK3	RAD21	SMO	WT1
BCORL1	CSF1R	FANCE	HIST1H3C	MAP3K1	NUP93	RAD50	SOCS1	XPO1
BIRC3	CSF3R	FANCF	HNF1A	MAP3K4	PALB2	RAD51	SOX2	XRCC2
BLM	CTCF	FANCG	HOXB13	MAP3K13	PARP1	RAD51B	SOX9	XRCC3
BMPR1A	CTLA4	FANCI	HRAS	MAPK1	PAX5	RAD51C	SOX17	ZBTB20
BRAF*	CTNNA1	FANCL	HUWE1	MAX	PAX8	RAD51D	SPEN	ZFHX3
BRCA1*	CTNNB1*	FAT1	ID3	MCL1	PBRM1	RAD52	SPOP	ZMYM2
BRCA2*	CUL3	FBXW7	IDH1	MDM2	PDCD1 (PD-1)	RAD54L	SPTA1	ZMYM3
BRIP1	CUX1	FGF1	IDH2	MDM4	PDCD1LG2 (PD-L2)	RAF1	SPTAN1	ZNF750
BTG1	CXCR4	FGF2	IGF1R	MECOM	PDGFRA*	RARA	SRC	ZRSR2
BTG2	CYSLTR2	FGF3	IKBKE	MED12	PDGFRB	RASA1	SRSF2	
BTK	DAXX	FGF4	IKZF1	MEF2B	PGR	RB1	STAG1	

<i>CACNA1A</i>	<i>DCK</i>	<i>FGF7</i>	<i>IL6ST</i>	<i>MEN1</i>	<i>PHF6</i>	<i>RBBP6</i>	<i>STAG2</i>
<i>CALR*</i>	<i>DDR2</i>	<i>FGF8</i>	<i>IL7R</i>	<i>MET*</i>	<i>PIK3C2B</i>	<i>RBM10</i>	<i>STAT3</i>
<i>CARD11</i>	<i>DDX3X</i>	<i>FGF9</i>	<i>INPP4B</i>	<i>MGA</i>	<i>PIK3C2G</i>	<i>RECQL4</i>	<i>STAT5B</i>
<i>CASP8</i>	<i>DDX41</i>	<i>FGF19</i>	<i>INPPL1</i>	<i>MGMT</i>	<i>PIK3CA</i>	<i>REL</i>	<i>STAT6</i>

†Contact [archer-tech@idtdna.com](mailto:archer-tech@idtdna.com) for the panel target file to view complete list of targeted regions.

\*Indicates that select regions of this gene are enabled for Structural Variant analysis in the GTF.

## SNP sites targeted for sample tracking

rs560681	rs430046	rs987640	rs10776839	rs12393891
rs740598	rs8078417	rs6444724	rs6530357	chrX:4429309
rs1498553	rs9951171	rs6811238	rs5971553	chrX:11314433
rs10773760	rs576261	rs13182883	rs5953060	chrY:6738552
rs1058083	rs1109037	rs214955	rs6524626	chrY:19490214
rs4530059	rs1523537	rs321198	rs5940270	
rs1821380	rs221956	rs4606077	rs722847	

SNPs may be used in combination to uniquely tag and track samples over time. Contact [archer-tech@idtdna.com](mailto:archer-tech@idtdna.com) for further details.

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## Revision History

Document Number	Date	Description of change
RA-DOC-481/REV01	August 2024	Initial release.